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SHEEP'S MILK CHEESES AS A SOURCE OF BIOACTIVE COMPOUNDS

- Review -

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Abstract: Since ancient times, sheep's milk cheeses have been a part of a human diet. Currently, their consumption is of great interest due to its nutritional and health values. The aim of the article was to review the chemical composition of sheep's milk cheeses and its main bioactive ingredients in the context of nutritional and health values. Sheep's milk cheeses are rich in functionally and physiologically active compounds such as: vitamins, minerals, fatty acids, terpenes, sialic acid, orotic acid and L-carnitine, which are largely originate from milk. Fermentation and maturation process additionally enrich them in other bioactive substances as: bioactive peptides, γ -aminobutyric acid (GABA) or biogenic amines. Studies show that sheep's milk cheese consumption may be helpful in the prevention of civilization diseases, i.e. hypertension, obesity or cancer. However, due to the presence of biogenic amines, people with metabolic disorders should be careful of their intake.

Keywords: biogenic amines, bioactive peptides, CLA, GABA, rumenic acid

INTRODUCTION

Cheeses are one of the oldest forms of food in the world. Their production is a natural way of preserving milk. At the same time, thanks to the activity of many beneficial microorganisms, products with an increased or modified nutritional value with the desired sensory characteristics are obtained, thereby increasing the diversity of food.

The start of cheese production is connected to taming of domestic animals, mainly sheep and goats. It is believed that the first cheeses were produced in the Middle East around 10,000 years ago in the Neolithic period (Walther et al., 2008). However, the oldest known evidence for the cheese production comes from about 5,500 BC, from today's Kujawy region, Poland (Salque et al., 2013). Despite the long world tradition of obtaining milk from sheep, cow's milk and dairy food produced of it dominate the world. Regarding to cheese, in 2018 (no newer data available) the world production of sheep's milk cheese was 726,421 tons, which constitute only ~4% of cow's milk cheese production (FAOSTAT, 2021). This is mainly the results from the marginal world production of sheep's milk as well as its regional significance. It

Received: 14.06.2021 Accepted in revised form: 25.10.2021 is also related to the seasonality of ewe's milking and their relatively small individual milk yield (Ptasińska-Marcinkiewicz, 2014).

As statistics show, the highest cheese consumption is currently in Europe, North America and Oceania. In 2018-2020, the average cheese consumption (expressed in kg per capita) for European Union was 20.9, United States 17.4, Canada 13.7, Australia 12.5, while for the continents of Africa and Asia it was only 0.7 and 0.6, respectively (OECD/FAO, 2021). Nevertheless, these data apply to all types of cheeses. There are no statistics available on the consumption of sheep's milk cheeses only. However, taking into account the sheep's milk cheeses supply quantity in 2018 (no newer data available), only in Greece it reached 36 grams per capita per day, while in other countries included in the database, it was only 0-5 grams per capita per day (FAOSTAT, 2021). The same statistics for cow's milk cheeses have higher and more varied values (0-96 g/capita/day), so it can be assumed that sheep's milk cheeses consumption is relatively low. It may be related to their availability (seasonality, distribution) and prices. Sheep's milk cheeses are often traditional, artisanal cheeses, and many of them are legally protected with a certificate of authenticity. The awareness of today's consumers (mainly from developed countries) about nutrition

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and its impact on health, increases the demand for good quality "healthy foods" (Balthazar et al. 2017). Sheep's milk cheeses, due to their properties, could fill this request.

As the composition and role of the individual components of sheep's milk cheeses were exploring, it was noted that some of them, in addition to nutrition of the human body, may have effects on its functioning (both, positive and negative). They are called bioactive compounds. According to Biesalski et al. (2009), these are "essential and non-essential compounds that occur in nature, are part of the food chain, and can be shown to have an effect on human health". The

BIOACTIVE COMPOUNDS IN SHEEP'S MILK CHEESES

Figure 1 presents the bioactive compounds from sheep's milk cheeses described in this article.

Minerals and vitamins

There are about twenty minerals (macro- and microelements) considered as essential for human. They play the structural, biochemical and nutritional functions in human body, so their adequate intake is necessary for optimal mental and physical health (Zamberlin et al., 2012).

The concentration of minerals in cheeses depend on the raw material (which is affected by sheep breed, the stage of lactation, environmental conditions, pasture type, soil contamination), cheese manufacturing (type of process, contaminations from equipment) and ripening (possible migration of minerals in cheese layers) (González-Martín et al., 2011). Also the type of rennet could affect the mineral content of cheese – animal rennet retains more zinc and phosphorus in cheese (Sanjuán et al. 1998).

Borys et al. (2006), conducted a research on the content of some minerals in sheep's milk and the degree of their retention in different types of cheeses made from it. They showed that sheep's milk cheeses are richer in calcium, phosphorus and magnesium than cow's milk cheeses. This is due to the fact that these minerals in sheep's milk are associated with casein micelles fraction forming the cheese curd. However, a part of them may also be present in a soluble, free form in milk. For this reason, cheeses contain several times more calcium and phosphorus than milk (fresh cheeses 4-5x, semihard cheeses 7-8x, hard cheeses 10x) (Chia et

sheep's milk is naturally rich in bioactive substances such as: vitamins, minerals, orotic acid, L-carnitine and a favourable profile of fatty acids. Nevertheless, some of substances are lost along with the whey during cheesemaking (Robinson and Tamime, 1996). The cheeses are also enriched with biologically active substances in the process of fermentation and maturation, by the breakdown of the main milk components – fats and proteins (Walther et al., 2008).

The aim of the article is to review the chemical composition of sheep's milk cheeses and its main bioactive ingredients in the context of nutritional and health values.

al., 2017). Optimal calcium, phosphorus and magnesium macroelements intake are important to prevent cardiovascular disease, osteoporosis, hypertension, insulin resistance and type 2 diabetes occurrence (Malara et al., 2017). In addition, it may prevent the formation of tooth decay (Rashidinejad et al., 2017).

Minerals such as sodium, potassium and chlorine are almost completely soluble in milk and are lost along the whey during cheesemaking (Chia et al., 2017). Nevertheless, cheeses can be a good source of these minerals due to the addition of salt. In European Union countries, cheese contribute around 10% of salt intake. Sodium is an essential for normal cell function, however, its high intake could contribute to high blood pressure and consequently increase the risk of cardiovascular and kidney diseases (European Commission, 2012). For this reason, it is recommended to limit its consumption. Cheeses with reduced sodium content is develop by decreasing NaCl levels or its substitution of with KCl, MgCl₂ and CaCl₂ (Cruz et al., 2011).

The consumption of sheep's milk cheeses complements the body's need for fat-soluble vitamins, mainly A (retinol) and E (α -tocopherol). Their amount depends on their content in milk (which is mostly related with the sheep's diet, but also farm practices or mastitis) and the cheesemaking process (Milewski et al., 2016; Rashidinejad et al., 2017). In addition, along with the time of cheese ripening their amount may decrease (Revilla et al., 2014; Gutiérrez-Peña et al., 2021). In regard to water-soluble vitamins, e.g. riboflavin (B_2) , niacin (B_{12}) or folate, their content in cheeses is lower than in milk. However, some B vitamins can be synthesized by microorganisms during the cheese ripening, which compensates for their loss during the cheese-making process (López-Expósito et al., 2017).



Figure 1. Schematic presentation of bioactive compounds from sheep's milk cheeses described in the article

According to Milewski et al. (2016) hard ripened rennet cheeses from sheep's milk, originating in Warmia-Masuria region in Poland, contain 2.6 μ g/g of vitamin A and 2.41 μ g/g of vitamin E. In the Italian Parmigiano sheep's milk cheese, these values range 0.86-6.83 μ g/g of vitamin A and 1.35-3.2 μ g/g of vitamin E (Perretti et al., 2004).

These two vitamins are essential for human. They can play diverse roles, however, eating food rich in these vitamins reduce the risk of occurrence cancers, can delay the body's aging (thanks to the antioxidant activity), as well as stimulate the immune system (Oruch and Pryme, 2012; Rizvi et al., 2014).

Orotic acid

Orotic acid (OA), formerly known as vitamin B_{13} , present in high level in ruminant milk, can have also a great influence on human body. In sheep's milk its amount varies from 0.45 to 1.95 mg/100 mL (Gajos and Krezlewicz, 1974). In cheeses, its quantity depends on the amount of soluble whey solids, degree of fermentation and time of cheese ripening (Larson and Hegarty, 1979; Urbienė and Leskauskaitė, 2006). Additionally, Manolaki et al. (2006) shows that the OA content in sheep's milk Feta cheese drops drastically during the first 2 days of its maturation, which may indicate the assimilation of OA by microorganisms.

OA is an organic acid (6-carboxyuracil) formed from carbamoyl phosphate and aspartic acid, which is a precursor of nucleotide compounds (Milewski, 2006). It has been shown that OA increase metabolism of folic acid and vitamin B₁₂, improved the condition of hearts with hypertrophy (West et al. 2017) and increase the growth of *Lactobacillus* sp. (Wright et al., 1950). However, people with impaired nucleotide metabolism can suffer from orotic aciduria (caused by a buildup of OA). In addition, studies on rats indicate that a diet rich in OA reduce Low Density Lipoprotein (LDL) cholesterol level in blood, but at the same time fat accumulating in the liver lead to its damage (Creasy et al., 1961; Živný et al., 2007).

Fatty acids

The most important and best studied bioactive ingredients of sheep's milk cheeses are fatty acids. Their amount depends primarily on the content in unprocessed milk (Nudda et al., 2005; Fernández et al., 2015). It has been shown that depending on the season and lactation period, the fatty acid profile of milk (Bielińska-Nowak and Czyżak-Runowska, 2016; Balthazar et al. 2017) as well as sheep's milk cheeses (Nudda et al., 2005; Kawecka and Sosin-Bzducha, 2014) change. In summer, saturated fatty acids are reduced in favor of unsaturated fatty acids (Walther et al., 2008). These changes are mainly caused by the way of animals feeding - with different availability of pastures and fatty acid composition of grasses (Nudda et al., 2005). The profile of free fatty acids in cheese depends also on the pasteurization process, the type of coagulant used and the parameters of cheese production (Kawecka and Sosin-Bzducha, 2014).

Studies show that sheep's milk cheeses are naturally rich in bioactive lipids (Table 1). The main fatty acids of Italian Gran Ovino cheese are C16:0 palmitic acid, C18:1 n-9 oleic acid and C14:0 myristic acid (Gaglio et al., 2019), while in Spanish artisanal hard cheeses – C16:0 palmitic and also C18:1 n-9 oleic acids (Estrada et al., 2019). Polish Bundz cheese contains large amounts of C10:0 capric acid, C16:0 palmitic acid, C18:1 n-9 oleic acid (Inoleic acid C18:2 n-6, conjugated linoleic acid (CLA) and linolenic acid C18:3 n-3 (Bonczar et al. 2009). Aguilar et al. (2014) showed that Chilean commercial sheep's milk cheeses were richer in C18:1 trans isomers (including vaccenic acid) compared to cow's and goat's milk cheeses, and had a lower thrombogenicity index.

Saturated fatty acids (SFA) constitute 60% to 70% of fatty acids in milk of ruminants and thus are the most abundant group of fatty acids in animal products (Markiewicz-Kęszycka et al., 2013). Depending on the length of the chain, we divide them into Short-Chain Fatty Acids (SCFA), Medium-Chain Fatty Acids (MCFA), Long-Chain Fatty Acids (LCFA) and Very Long-Chain Fatty Acids (VLCFA). The presence of saturated SCFA (1-6 carbon atoms in the chain) and MCFA (6-12 carbon atoms) is a unique feature of milk fat. In ruminants, their synthesis takes place de novo in the mammary gland (Rutkowska et al., 2015). They are absorbed from the gastrointestinal tract without bile acids action, quickly penetrating into the blood without esterification (Cichosz and Czeczot, 2012b). Experimental studies show that MCFAs in the diet suppress the deposition of fat due to improved thermogenesis and fat oxidation in both. animals and humans (Nagao and Yanagita, 2010).

Monounsaturated fatty acids (MUFA) constitute 20%-35% of sheep's milk fat and includes e.g. vaccenic acid (C18:1 cis-7) (Markiewicz-Kęszycka et al., 2013). However, the trans isomer of vaccenic acid (C18:1 trans-11) is mostly present in sheep's milk cheeses. It is synthesized in the rumen of animals by bacteria and is a precursor of CLA (Rutkowska et al., 2015). It does not show proatherosclerotic activity, which is attributed to trans fatty acids. Contrary, studies in animal models show that a vaccenic acid-rich diet (C18:1 trans-11) has a hypolipidemic effect, which may prevents the formation of atherosclerotic lesions (Bassett et al., 2010) and alleviate the features of the metabolic syndrome (Jacome-Sosa et al., 2014). In addition, it has been shown that both isomers, cis and trans, exhibit anti-carcinogenic effects (Banni et al., 2001; Miller et al., 2003; Sauer et al., 2004; Lim et al., 2014).

The group of polyunsaturated fatty acids (PUFA) with health-promoting properties contained in sheep's milk cheeses include mainly omega-3,-6,-9, CLA, as well as eicosapentaenoic acid (EPA) and

docosahexaenoic acid (DHA). It has been shown that increasing the consumption of essential PUFAs, including linoleic acid, α -linolenic acid, EPA, DHA and γ -linolenic acid, reduces the risk of diet-related diseases (Janczy, 2012).

The most important and best-studied bioactive fatty acid found in sheep's milk cheeses is the so-called rumenic acid - conjugated dienes of C18:2 linoleic acid (Conjugated Linoleic Acid, CLA). CLA arises mainly in the mammary glands of animals as a result of the Δ 9-desaturase of vaccenic acid and partly through the bioconversion of PUFA by anaerobic bacteria that takes place in the rumen (Recio et al., 2009; Cichosz and Czeczot, 2012a). Accordingly, rumenic acid is found only in the meat and milk of ruminants, including sheep. Numerous studies have confirmed its health-promoting effects. Currently market offers many dietary supplements using these properties. Their main task is to support weight loss by inhibiting enzymes responsible for the deposition of adipose tissue (Cichosz and Czeczot, 2012a).

Animal studies have shown that enrichment of diet in CLA increases the body's immunity and improves the metabolism of cholesterol and triacylglycerol in the blood plasma, thereby inhibiting the development and pathogenesis of atherosclerosis (Pariza, 1999; Roche et al., 2001). In addition, a small (1%) dose of this fat in the diet has an anticancer effect, causing inhibition of carcinogenesis at various stages of its development (Kowalska and Cichosz, 2013). This is mainly related to the antioxidant activity of CLA (Cichosz and Czeczot, 2012a).

Until recently, nutritionists have not recommended frequent consumption of cheese and animal products, mainly due to the high content of sodium, saturated fat and trans fatty acids. Consumption of dairy products was associated with an increased risk of obesity, atherosclerosis and cancer (Krzęcio-Nieczyporuk and Antosik, 2015). However, these assumptions are not confirmed by the latest research. Sofi et al. (2010) conducted preliminary studies that has been shown that the short-term (10weeks) consumption of regional Italian hard Pecorino sheep's milk cheese, naturally rich in CLA cis and trans isomerism, significantly improves the value of anti-atherosclerotic markers. Additionally, Tong et al. (2017) showed that long-term consumption of cheese is not associated with an increased risk of death, as previously thought. Moreover, in countries with the highest consumption of ripening cheeses rich in CLA (France, Italy, Greece), a much lower mortality rate was observed due to, among others breast cancer (Kowalska and Cichosz, 2013).

| Table 1. Weall values of fatty | acias in vario | us sheep s min | x enceses | | | |
|--------------------------------|----------------|----------------|-----------|------|---------|------------------------------------|
| Type of cheese | SFA | MCFA | MUFA | PUFA | CLA | Reference |
| | | | [g/100g] | | | - |
| Gran Ovino | 68.40 | no data | 22.97 | 8.62 | 1.05 | Gaglio et al. (2019) |
| Pecorino Carmasciano | 51.30 | no data | 19.80 | 4.30 | 0.52 | Marrone et al. (2014) |
| Pecorino di Farindola | 74.62 | 41.23 | 21.25 | 4.16 | 1.02 | Schirone et al. (2011) |
| Pecorino | 67.69 | 21.70 | 26.83 | 5.48 | 0.78 | Prandini et al. (2007) |
| Pecorino | 65.94 | 22.27 | 23.80 | 4.93 | 0.11 | Prandini et al. (2011) |
| Roquefort | 70.10 | 26.12 | 21.11 | 3.92 | 0.88 | |
| Semi-hard ripened | 66.32 | 13.40 | 28.47 | 4.60 | 0.84 | Jarzynowska and Kłopotek (2013) |
| Hard ripened | 74.07 | no data | 22.04 | 3.90 | 1.09 | Milewski et al. (2016) |
| Feta | 70.20 | no data | 21.00 | 4.7 | 0.18 | Zlatanos et al. (2002) |
| Kefalotyri | 71.69 | no data | 23.33 | 5.00 | 1.09 | Govari et al. (2020) |
| (made in December) | | | | | | _ |
| Kefalotyri | 69.20 | no data | 24.78 | 6.04 | 1.48 | |
| (made in April) | | | | | | |
| Serra da Estrela | 73.4 | no data | 20.3 | 6.3 | no data | Lima et al. (2019) |
| Azeitão | 73.8 | no data | 17.5 | 4.11 | 0.8 | Partidário et al. (2008) |
| Évora | 71.2 | no data | 19.4 | 4.35 | 0.8 | _ |
| Nisa | 67.9 | no data | 22.1 | 4.60 | 1.0 | |
| Roja Mallorquina | 21.69 | 13.73 | 6.52 | 1.99 | 0.14 | Gutiérrez-Peña et al. |
| (fresh) | | | | | | (2021) |
| Roja Mallorquina | 14.87 | 9.12 | 3.60 | 1.14 | 0.21 | |
| (ripened) | | | | | | |

Table 1. Mean values of fatty acids in various sheep's milk cheeses

SCFA – Saturated Fatty Acid, MCFA – Medium-Chain Fatty Acids; MUFA – Monounsaturated Fatty Acids; PUFA – Polyunsaturated Fatty Acids, CLA – Conjugated Linoleic Acids

The recent discoveries of the beneficial effects of cheese consumption on human health could be related with changes in milk composition over the years (different breeding systems and animal nutrition), greater knowledge of bioactive compounds, as well as improved analytical methods (Raynal-Ljutovac et al., 2008).

It is known that the fatty acid profile of sheep's milk could be modified to a greater extent than that of cow's milk (by changing animal feeding). Consequently, the creation of functional sheep's milk cheeses is simpler and in this respect, eating them may have a greater impact on our health. This is confirmed by recent studies. Pintus et al. (2013) investigated the effects of eating Italian Pecorino cheese, naturally enriched in linoleic acid (by feeding sheep's with extruded linseed), on the health of people with diagnosed mild hypercholesterolemia. For this purpose, 42 adult volunteers were eating control or enriched cheese (90 g/day), for 3 weeks. It has been proven that, in opposite to control cheese, intake of naturally enriched cheese significantly increased the plasma concentration of CLA, vaccenic acid, the n-3 fatty acids Alpha Lipoic Acid (ALA) and EPA and decreased endocannabinoid anandamide and LDLcholesterol. Similar research was done by Murru et al. (2018). 15 adult volunteers ate ALA and CLA enriched sheep's milk cheese (90 g/day) for 4 weeks. It resulted increasing the EPA (80%) and DHA (20%)plasma level. higher CLA incorporation (240%), as well as improving the n-3 highly unsaturated fatty acids (HUFA) score (28%). Further research consisted in administering to 36 volunteers variously enriched cheeses made from sheep's, cow's or goat's milk (50 g/day) for 2 months. Consumption of each of the cheese improved the n-3 HUFA score by increasing the DHA plasma level and the effect was proportional to their CLA content.

Terpenes

Terpenes are isoprene-based compounds, mainly of plant origin. Therefore, their amount and composition in milk and cheese is mainly determined by their content in the diet of animals. The milk pasteurization and cheesemaking process could also affect the terpene profile (Cornu et al. 2005). In addition, terpenes may undergo microbial bioconversion (Mikami, 1988). As the qualitative and quantitative composition of terpenes in plants is influenced by such factors as: their species, growth stage, soil, climate, geographical location and the management of grassland (Poulopoulou et al., 2011), terpenes in cheeses are investigated for their potential function as biomarkers of the cheese origin according to the way the animals are fed (Favaro et al., 2005). In this way it was possible to distinguish the highland from lowland cheeses (Valdivielso, et al. 2017; Moran et al., 2019). However, Poulopoulou et al. (2011) showed that terpenes as cheese biomarkers may not be reliable.

Various terpenes have been found in sheep's milk cheeses (Table 2). Limonene and α -pinene were most frequently detected. Depending on the amount of terpenes in the product, they could affect the sensory properties of cheese, although their importance in creating the flavor of the cheese appears to be controversial (Curioni and Bosset, 2002).

Terpenes are known for their broad therapeutic properties. However, in the context of their presence in cheeses, it has not yet been investigated whether they have an impact on the health of the consumer.

| Type of cheese | Terpene | Reference |
|-----------------------------------------|------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------|-------------------------------|
| Halloumi | α-pinene | Papademas and Robinson (2002) |
| | β-pinene | |
| | copaene | |
| | thymol | |
| | α-caryophyllene | |
| | β-caryophyllene | |
| | δ-cadinene | |
| Manchego | D-limonene | Barron et al. (2005) |
| <u> </u> | cymene | |
| Idiazabal | α-pinene | Barron et al. (2007) |
| | <i>D</i> -limonene | |
| Vastedda della valle del Belìce | Terpene a -pineneF β -pineneF $copaenefthymola-caryophyllene\beta-caryophyllene\delta-cadineneD-limonenecymenea-pineneD-limonenea-pinenef\beta-pinenef\beta-pinenef\beta-pinenef\beta-pinenef\beta-pinenef\beta-pinenef\beta-pinenef\beta-pinenef\beta-pinenef\beta-pinenef\beta-pinenef\beta-pinenef\beta-pinenef\beta-pinenef\beta-pinenef\beta-pinenef\beta-pinenef\beta-pinenef\beta-pinenef\beta-pinenef\beta-pinenef\beta-pinenef\beta-pinenef\beta-pinenef\beta-pinenef\beta-pinenef\beta-pinenef\beta-pinenef\beta-pinenef\beta-pinenef\beta-pinenef\beta-pinenef\beta-pinenef\beta-pinenef\beta-pinenef\beta-pinenef\beta-pinenef\beta-pinenef\beta-pinenef\beta-pinenef\beta-pinenef\beta-pinenef\beta-pinenef\beta-pinenef$ | Verzera et al. (2010) |
| | β-pinene | |
| | limonene | |
| | <i>o</i> -cymene | |
| | caryophyllene | |
| Gran Ovino | D-limonene | Gaglio et al. (2019) |
| Urfa | α-pinene | Atasoy et al. (2013) |
| | β-pinene | |
| | camphene | |
| | β-myrcene | |
| | limonene | |
| | <i>o</i> -cymene | |
| Piacentinu Ennese | ocimene | Horne et al. (2005) |
| | terpinolene | |
| | rose oxide | |
| | L-carvone | |
| | citronellol | |
| | a-terpineol | |
| | 1 | |
| Table 3. The content of GABA in various | sheep's milk cheeses | |
| Type of cheese | GABA[mg/kg] | Reference |
| Pecorino | 289 - 391 | Siragusa et al. (2007) |
| Pecorino di Farindola | 554 - 672 | Tofalo et al. (2019) |
| Spanish artisinal | 100 - 980 | Diana et al. (2014) |
| Fruhe | 28.8 | Murgia et al. (2020) |
| Roncal | 271 - 986 | Muñoz et al. (2003) |
| Roncal type | 33.7 - 891 | Irigoven et al. (2007) |

238.1 - 1638.9

Table 2. Terpenes found in various sheep's milk cheeses

Serra da Estrela

Tavaria et al. (2003)

γ-Aminobutyric acid

 γ -Aminobutyric acid (GABA) is a non-protein amino acid, which presence in food is recently sought after due to its potential health benefits. The main source of GABA in cheese are Lactic Acid Bacteria with glutamic acid decarboxylase enzyme activity, mainly *Lactobacillus* spp. During the fermentation and proteolysis, a large amount of L-glutamate, a precursor of GABA, is released from native caseins (Tofalo et al., 2019). For this reason, except microbiota, an amount of protein in raw milk, type of coagulant used, as well as time of ripening affect this amino acid content in cheese (Santiago-López et al., 2018). In addition, research showed that with the decreases of cheese pH, the amount of GABA increases (Nomura et al., 1998).

The GABA content in cheese correlates with the increased number of cheese eyes, although it has no direct or indirect effect on its flavor (Estrada et al., 2019). The concentration of GABA in some sheep's milk cheeses was examined (Table 3) and it is very variable.

In humans, GABA is an inhibitory neurotransmitter that can perform various functions in the central and peripheral nervous system as well as in some nonneuronal tissues (Watanabe et al., 2002). It was proved that daily consumption of GABA-containing dairy product could be helpful in regulating and stabilizing blood pressure of people with high normal pressure and mild hypertension (Inoue et al., 2003; Kajimoto et al., 2004). Furthermore, researches indicate that GABA intake could alleviate human stress and anxiety, while enhancing its immune responses (Abdou et al., 2006; Yoto et al., 2012). Due to the positive impact of GABA consumption on human health, attempts are being made to create functional GABA-enriched dairy foods, including cheese, using GABA-producing microorganisms (Linares et al., 2016; Carafa et al., 2019; Redruello et al. 2021).

L-carnitine

L-carnitine $(L-\beta-hydroxy-\gamma-N,N,N-trimethyl$ aminobutyric acid) is an essential cofactor of fattyacid metabolism, endogenously biosynthesizedfrom the amino acids L-lysine and L-methionine orobtained from food sources (Goa et al., 1987). Thedaily requirement of L-carnitine in humans rangesbetween 2 and 12 µmol/kg body weight/day (0.3 and1.9 mg/kg/day) and may even be higher for athletesand pregnant women (Demarquoy et al., 2004).

According to Bodkowski et al. (2011), sheep's milk contains a lot of L-carnitine amino acid – on average 11.08 mg/100 mL, about 32.1% more than cow's milk. The high level of carnitine in sheep's milk may reflect the specific fatty acid utilization needs of the newborn animal (Woollard et al.,1999). Unlike milk, sheep's milk cheeses are its less-rich source. Due to its hydrophilicity, it is lost along with the whey during cheese production and along with the loss of water during ripening (Bodkowski et al., 2011). Nevertheless, sheep's milk cheeses contain about 50% more L-carnitine than cow's milk cheeses (Seline and Johein, 2007).

L-carnitine is widely known as a supplement supporting slimming. It transports chains of fatty acids into the mitochondrial matrix, thus enabling cells to break down and draw energy from stored fat stores, thereby helping to lose weight and detoxify cells (Pękala et al., 2011). In addition, due to its antioxidant properties, it has a potential anticancer effect. Increased levels of other antioxidants by L-carnitine were also demonstrated, as well as its ability to chelate iron, cadmium or lead (Rospond and Chłopicka, 2013).

Bioactive peptides

Bioactive peptides are short chains containing 2-20 amino acid residues, which are released as a result of proteolysis and gained new pro-healthy properties (Balthazar et al., 2017). In cheese, their source is mainly casein broken down by milk enzymes, rennet, starter cultures or secondary microbiota (López-Expósito et al., 2017). However, peptides released during bioactive cheese production may become inactive after proteolysis during its long ripening periods (Alhaj and Kanekanian, 2014). It makes their presence dependent on the type and properties of cheese (Barać et al., 2017).

Some in vitro and in vivo studies were carried out to determine the properties of bioactive peptides and their impact on the human body. It has been proven that some active peptides derived from sheep's milk case in fragments: α s1-, α s2- and β - have antimicrobial properties, e.g. on Escherichia coli (Benkerroum, 2010). They can also exhibit strong antioxidant activity (Barać et al., 2017). Others, obtained from the decomposition of k-casein and lactoferrin, inhibit thrombin-induced platelet aggregation, thus exhibiting anticoagulant effects (Marcone et al., 2017). Some peptides derived from case β inhibit the angiotensin converting enzyme I (ACE), thereby contributing to lowering blood pressure (Balthazar et al., 2017; Zdrojewicz et al., 2017). However, there is no conclusive evidence that bioactive peptides present in food, other than di- and tripeptides, can penetrate the intestinal wall hepatic and enter the portal system in physiologically relevant concentrations (Miner-Williams et al., 2014).

Pisanu et al. (2015) compared the composition of bioactive peptides in ripening sheep's milk cheeses made from raw and pasteurized milk. They showed, that there are significant differences in the peptide profiles of the examined cheeses in terms of individual sequences and their relative amounts. In sillico analysis of the properties of bioactive peptides (BIOPEP software) showed that 69% of the identified bioactive peptides have unknown properties and the rest are likely to have immunomodulatory (11%), ACE inhibitor (8%), antibacterial (5%), antioxidant (4%), anticancer and opioid-agonistic (1%) properties. (2%)Significantly higher content of peptides with immunomodulatory properties and ACE inhibitors was found in raw milk cheese and opioidantagonistic activity in pasteurized milk cheeses.

The biological activity of peptides, due to the complexity of research, is mostly demonstrated in vitro (Table 4), which does not always reflect the effects obtained in a living organism (Darewicz et al., 2015). Further research is needed to better understand their impact on human health.

Sialic acid

Sheep's milk is particularly rich in a sialic acid (Sia) – family of 9-carbon acidic sugars contained in oligosaccharides, glycolipids and glycoproteins (Nakano et al., 2001). Its amount in milk depends on genetics, breed of animals, their nutrition, geographical location, lactation period or a combination of these factors (Karunanithi et al., 2013). During cheesemaking, by the action of rennet, κ -casein from milk is hydrolyzed to para- κ -casein, which remains in the curd, and glycomacropeptide, which is removed with the whey with all the κ -casein sugars, including Sia (Costa et al., 2019). In literature, there is no data about its levels in sheep's milk or cheeses.

Sia occurs mainly as N-acetylneuraminic acid (Neu5Ac) and N-glycolylneuraminic acid (Neu5Gc). Neu5Ac, naturally present in human milk and body, plays an important role in infant brain development and cognition (Wang, 2009).

Table 4. Bioactive peptides derived from different types of sheep's milk cheeses and their functional effects demonstrated in vitro

| Type of cheese | Sequence of bioactive peptide CN fragment | | Functional effect | Reference |
|------------------------|-------------------------------------------|--------------------------------|------------------------------|------------------------|
| Pecorino Romano | GLSPEVLNENLL | α _{S1} -CN f(10–21) | antibacterial properties | Rizzello et al. (2005) |
| Romano | RFVVAPFPE | α _{S1} -CN f(22–30) | and Gram negative | (2003) |
| | VVAPFPEV | α _{S1} -CN f(24–31) | Dacterna | |
| | VMFPPQSVL | β-CN f(155–163) | _ | |
| Canestrato Pugliese | MPIQAF | β-CN f(183–188) | | |
| Idiazábel | QP, FP | various fragments | angiotensin- Gómez-Ru | |
| | DKIHP | β-CN f(47–51) | (ACE)-inhibitory | et ul. (2000) |
| | DKIHPF | β-CN f(47–52) | activities | |
| Roncal | QP, PP | various fragments | _ | |
| | РКНР | α _{S1} -CN f(2–5) | | |
| | DKIHP | β-CN f(47–51) | | |
| | DKIHPF | β-CN f(47–52) | _ | |
| Cabrales | PP | various fragments | | |
| | DKIHP | β-CN f(47–51) | | |
| | DKIHPF | β-CN f(47–52) | | |
| Roquefort- type | TDAPSFSDIPNPIGSENSGK | α _{S1} -CN f(189–208) | antioxidant and angiotensin- | Meira et al. (2012) |
| -9F- | DIPNPIGSENSGKTTMPLW | α _{S1} -CN f(196–214) | converting-enzyme | (===) |

| | IPNPIGSENSGKIT | α _{S1} -CN f(197–210) | (ACE)-inhibitory | | | |
|----------|-----------------------|---------------------------------|-------------------|---------------|--|--|
| | YQGPIVLNPWDQVK | α ₈₂ -CN f(116–129) | properties | | | |
| | YQGPIVLNPWDQVKR | α ₈₂ -CN f(116–130) | | | | |
| | GPIVLNPWDQVKR | α _{s2} -CN f(118–130) | | | | |
| | VLNPWDQVKR | α _{s2} -CN f(121–130) | | | | |
| | NAGPFTPTVNR | α _{s2} -CN f(131–141) | | | | |
| | KEMPFPKYPVE | β-CN f(122–132) | | | | |
| | WMHQPPQPLPPTVMFPPQSVL | β-CN f(158–178) | | | | |
| | MHQPPQPLPPTVMFPPQSVL | β-CN f(159–178) | | | | |
| | HQPPQPLPPTVMFPPQSVL | β-CN f(160–178) | | | | |
| | YQEPVLGPVRGPFPI | β-CN f(206–220) | | | | |
| | OEPVLGPVRGPFPI | β-CN f(207–220) | | | | |
| | OEPVL GPVR GPFPIL V | β-CN f(207–222) | | | | |
| | PVI GPVRGPFPI | $\beta = CN f(209 - 222)$ | | | | |
| | | $\beta = CN f(211, 220)$ | | | | |
| | | p-CN I(211-220) | | | | |
| Manchego | VVAPFPE | α_{s1} -CN f(24-30) | converting-enzyme | Gómez-Ruiz | | |
| | FPE | α _{S1} -CN f(28-30) | (ACE)-inhibitory | et al. (2002) | | |
| | KQMK | α _{S1} -CN f(58-61) | activities | | | |
| | DVPSERY | α _{S1} -CN f(85-91) | | | | |
| | DVPSERYLG | α _{S1} -CN f(85-93) | | | | |
| | VPSERY | α _{S1} -CN f(86-91) | | | | |
| | VPSERYL | α ₈₁ -CN f(86-92) | | | | |
| | KKYNVPQ | α _{S1} -CN f(102-108) | | | | |
| | KKYNVPQL | α _{S1} -CN f(102-109) | | | | |
| | LEIVPK | α _{S1} -CN f(109-114) | | | | |
| | LKKISQ | α _{S2} -CN f(165-170) | | | | |
| | AWPQ | α ₈₂ -CN f(176-179) | | | | |
| | TQPKTNAIPY | α _{s2} -CN f(195-204) | | | | |
| | VRYL | α _{s2} -CN f(205-208) | | | | |
| | IPY | α _{Si2} -CN f(202-204) | | | | |
| | REQEEL | β-CN f(1-6) | | | | |
| | DKIHP | β-CN f(47-51) | | | | |
| | VPKVKE | β-CN f(95-100) | | | | |
| | VPKVKET | β-CN f(95-101) | | | | |
| | GPVRGPFP | β-CN f(197-204) | | | | |
| | VRGPFP | β-CN f(199-204) | | | | |
| | | , | | | | |

In contrast, the influence of Neu5Gc on human health is not vet clear. It is not produced in human body due to a deletion in the gene encoding the enzyme responsible for its formation - CMP-Neu5Ac hydroxylase (Karunanithi et al., 2013). However, Neu5Gc builds up in the body from food sources, such as red meat and dairy. The research of Samraj et al. (2015) indicates that long-term consumption of Neu5Gc (from red meet) could lead to chronic inflammation, resulting from the reaction of the human immune system. It can participate in the formation of cancerous changes. In addition, the long-term accumulation of this compound may result in the appearance and development of certain diseases in the elderly (Szczurek, 2004). However, more research should be done in this regard.

Biogenic amines

Biogenic amines (BA) are a low-molecular nitrogen compounds produced during the fermentation and ripening of cheeses, mainly as a result of the decarboxylation of amino acids by microorganisms. Therefore, the content of BA in cheese depends mainly on the type of microorganisms occurring during fermentation and maturation, but also on many other factors such as availability of substrates, temperature and cheese ripening / storage time, their pH, salt content, oxygen availability, redox potential, level of sugar, added herbs or used technological processes (Bonczar et al., 2017). For these reasons, the quantitative and qualitative composition of BA in sheep's milk cheeses is extremely variable, even for one type of cheese (Schirone et al., 2012). High concentration of BA in cheese could indicate poor hygienic conditions during its production and storage (Martuscelli et al., 2005).

Several studies have been done to check the level of BA in sheep's milk cheeses (Table 5). They confirm high variability. Researches show that the amount of BA in cheese increases with its ripening time, and pasteurization does not guarantee a reduction.

Table 5. The content of individual biogenic amines in various sheep's milk cheeses

| Table 5. The content of individual ofogenic annues in various sheep's finite cheeses | | | | | | | |
|--------------------------------------------------------------------------------------|-----------------|-----------------|------------|------------|-------------|--------------|----------------------------------------|
| Type of | PHE | PUT | CAD | HIS | TYR | total BA | Reference |
| cheese | | | [r | ng/kg] | | | |
| Pecorino di Farindola | 0.0–127.1 | 9.9–394.1 | 26.8–276.1 | 0.0–21.8 | 52.3–1171.3 | 209.0-1839.0 | Schirone et al. (2011) |
| Pecorino | nd-232.4 | nd-986.0 | nd-2127.6 | nd-761.4 | nd-1771.3 | 10.3–5860.6 | Schirone et al. (2013) |
| Pecorino | 24–144 | 22–512 | 2–262 | nd-23 | 147–1132 | 274–2161 | Torracca et al. (2015) |
| Manchego | nd-49.8 | nd-668.3 | 1.8-803.1 | 16.1–217.9 | 5.6-326.8 | 43.7-881.7 | Poveda et al. (2015) |
| Terrincho | 12.9–237.8 | 82.6-446.5 | 48.6–239.6 | nd-10.9 | nd-283.1 | 428.0–922.0 | Pintado et al. (2008) |
| Zamorano | 3–120 | 10–190 | 5–35 | 1–55 | 1-85 | no data | Combarros- Fuertes et al. (2015) |
| Feta | 0.77-7.04 | 1.6–193 | 0.2-82.8 | 0.0-84.6 | 0.0–246 | 12.2–617 | Valsamaki et al. (2000) |
| Fiore Sardo | 2–16 | nd-450 | 6–40 | 4–210 | 60–700 | 170–1100 | Zazzu et al. (2019) |
| Azeitão | no data | nd-137 | 161-260 | 414-818 | 72.3–445 | no data | Pinho et al. (2001) |
| blue- veined | 13.25– 61.44 | 17.28– 33.46 | no data | no data | 7.15–52.20 | no data | Calzada et al. (2013) |
| bryndza | no data | 22.1-60.9 | 16.5-42.6 | 24.2 | 34.6-107.4 | 73.2-222.2 | |
| smoked | no data | 16.2–99.9 | 62.6-80.7 | nd | 8.9–38.3 | 25.1-177.1 | _ |
| fresh | no data | 20.7 | 19.6 | nd | nd | nd-40.3 | |
| unripened | no data | 55.3-118.2 | 11.4–35.8 | nd | 10.2-11.1 | nd-140.3 | Buňková et al. (2013) |
| pasta filata type | no data | nd | nd | nd | nd | nd -13.2 | - (2013) |
| brined | no data | 229.5 | 125.6 | nd | 23.1-174.6 | 37.2–529.8 | _ |
| flavoured | no data | 108.8 | nd | nd | 114.7 | nd-223.5 | |
| | | | | | | | |

PHE-phenylethylamine; PUT - putrescine; CAD - cadaverine; HIS - histamine, TYR - tyramine; BA - biogenic amines; nd - not detected

BAs have an ambiguous effect on human health. They play an important role in the human body including as precursors in the synthesis of hormones, proteins, alkaloids or nucleic acids, as well as by participating in the regulation of blood pressure, allergic reactions or cell growth control (Jansen et al., 2003). However, consumption of a large amount of BA may have an adversely affect the health of consumers, especially people with metabolic disorders, causing symptoms such as nausea, headaches, rash, arrhythmia or changes in blood pressure (Santos, 1996).

Research to date indicates that healthy people not taking monoamine oxidase inhibitor (MAOI) drugs could be exposed to a level of 25-50 mg of histamine and 600 mg of tyramine per person per meal (EFSA, 2011). However, currently there are none specific toxic doses for individual BA in cheeses determined, as they depend on each

CONCLUSIONS

Sheep's milk cheeses contain many bioactive ingredients both derived from milk and formed during its fermentation and / or maturation, largely due to the action of microorganisms. For this reason, the content of this individual components is variable

individual's detoxification body mechanisms (Ordonez et al., 1997).

The level of BA in cheeses can be reduced by the use of high pressure processing, y-irradiation, applications of additives (like spices and herbs), decarboxylase reduction of activity and temperature, and the most popular way, use of a selected starter culture (Ercan et al., 2013). Renes et al. (2019) was managed to produce a sheep's milk cheese with reduced level of BA and at the same time a higher GABA content by using autochthonous Lactococcus lactis subsp. lactis and Lactococcus lactis subsp. cremoris as starters and Lactobacillus plantarum as adjunct culture. As sheep's milk cheeses are often produced in a traditional way, perhaps such an approach to cheese production could increase their safety, while preserving the native character and enhancing the health-promoting properties.

and depends mainly on the milk composition, type of cheese, time of ripening and microbiota involved in the process. Contrary to popular belief, sheep's milk cheese consumption has a proven positive effect on human nutrition and health. However, further long-term research is needed to assess its role as a functional food.

REFERENCES

- 1. Abdou A.M., Higashiguchi S., Horie K., Kim M., Hatta H. & Yokogoshi H. (2006). Relaxation and immunity enhancement effects of γ -Aminobutyric acid (GABA) administration in humans. *Biofactors*, 26(3), 201-208. DOI: 10.1002/biof.5520260305.
- 2. Aguilar C., Toro Mújica P., Vargas Bello E., Vera R.R., Ugalde C., Rodríguez S., Briones I. (2014). A comparative study of the fatty acid profiles in commercial sheep cheeses. *Grasas y aceites*, 65(4). DOI: 10.3989/gya.0460141.
- 3. Alhaj O.A. & Kanekanian A. (2014). Milk-derived bioactive components from fermentation. *Milk and Dairy Products as Functional Foods*, 237-288. DOI: 10.1002/9781118635056.ch8.
- 4. Atasoy A.F., Hayaloglu A.A., Kırmacı H., Levent O. & Türkoğlu H. (2013). Effects of partial substitution of caprine for ovine milk on the volatile compounds of fresh and mature Urfa cheeses. *Small Ruminant Research*, *115*(1-3), 113-123. DOI: 10.1016/j.smallrumres.2013.09.002.
- Balthazar C.F., Pimentel T.C., Ferrão L.L., Almada C.N., Santillo A., Albenzio M., Mollakhalili N., Mortazavian A.M., Nascimento J.S., Silva M.C., Freitas M.Q., Sant'Ana A.S., Granato D. & Cruz A.G. (2017). Sheep milk: Physicochemical characteristics and relevance for functional food development. *Comprehensive Reviews in Food Science and Food Safety*, 16(2), 247-262. DOI: 10.1111/1541-4337.12250.
- 6. Banni S., Angioni E., Murru E., Carta G., Paola Melis, M., Bauman, D., Dong Y. & Ip C. (2001). Vaccenic acid feeding increases tissue levels of conjugated linoleic acid and suppresses development of premalignant lesions in rat mammary gland. *Nutrition and cancer*, *41*(1-2), 91-97. DOI: 10.1080/01635581.2001.9680617.
- 7. Barać M., Pešić M., Vučić T., Vasić M. & Smiljanić M. (2017). White cheeses as a potential source of bioactive peptides. *Mljekarstvo*, 67(1), 3-16. DOI: 10.15567/mljekarstvo.2017.0101.
- Barron L.J.R., Redondo Y., Flanagan C.E., Pérez-Elortondo F.J., Albisu M., Nájera A.I., de Renobales M. & Fernández-García E. (2005). Comparison of the volatile composition and sensory characteristics of Spanish PDO cheeses manufactured from ewes' raw milk and animal rennet. *International Dairy Journal*, 15(4), 371-382. DOI: 10.1016/j.idairyj.2004.08.005.

- Barron, L. J. R., Redondo, Y., Aramburu, M., Gil, P., Pérez-Elortondo, F. J., Albisu, M., Nájera A.I., de Renobales M. & Fernández-García E. (2007). Volatile composition and sensory properties of industrially produced Idiazabal cheese. *International Dairy Journal*, 17(12), 1401-1414. DOI: 10.1016/j.idairyj.2007.04.001.
- Bassett C.M., Edel A.L., Patenaude A.F., McCullough R.S., Blackwood D.P., Chouinard P.Y., Paquin P., Lamarche B. & Pierce, G.N. (2010). Dietary vaccenic acid has antiatherogenic effects in LDLr-/mice. *The Journal of nutrition*, 140(1), 18-24. DOI: 10.3945/jn.109.105163.
- 11. Benkerroum N. (2010). Antimicrobial peptides generated from milk proteins: a survey and prospects for application in the food industry. A review. *International Journal of Dairy Technology*, *63*(3), 320-338. DOI: 10.1111/j.1471-0307.2010.00584.x.
- 12. Bielińska-Nowak S. & Czyżak-Runowska G. (2016). Jakość higieniczna, wydajność i podstawowy skład mleka owczego w zależności od fazy laktacji. *Roczniki Naukowe Polskiego Towarzystwa Zootechnicznego*, *12*(1), 9-15.
- Biesalski H.K., Dragsted L.O., Elmadfa I., Grossklaus R., Müller M., Schrenk D., Walter P. & Weber P. (2009). Bioactive compounds: Definition and assessment of activity. *Nutrition*, 25(11-12), 1202-1205. DOI: 10.1016/j.nut.2009.04.023.
- 14. Bodkowski R., Patkowska-Sokoła B., Nowakowski P., Jamroz D. & Janczak M. (2011). Produkty pochodzące od przeżuwaczy najważniejsze źródło L-karnityny w diecie człowieka. *Przegląd Hodowlany*, *10*, 22-25.
- 15. Bonczar G., Filipczak-Fiutak M., Pluta-Kubica A. & Duda I. (2017). Aminy biogenne w serach występowanie i zagrożenia. *Medycyna Weterynaryjna*, 73(3), 136-143.
- 16. Bonczar G., Regula-Sardat A., Pustkowiak H. & Zebrowska A. (2009). Wpływ substytucji mleka owczego mlekiem krowim na właściwości bundzu. Żywność Nauka Technologia Jakość, 5(66), 96-106.
- 17. Borys M., Pakulski T., Borys B., Pakulska E. & Węgrzyn E. (2006). The content and retention of some major and trace minerals in sheep's milk and cheese. *Archiv fur Tierzucht*, 49, 263-267.
- Buňková L., Adamcová G., Hudcová K., Velichová H., Pachlová V., Lorencová E. & Buňka F. (2013). Monitoring of biogenic amines in cheeses manufactured at small-scale farms and in fermented dairy products in the Czech Republic. *Food Chemistry*, 141(1), 548-551. DOI: 10.1016/j.foodchem.2013.03.036.
- 19. Calzada J., Del Olmo A., Picon A., Gaya P. & Nuñez M. (2013). Proteolysis and biogenic amine buildup in high-pressure treated ovine milk blue-veined cheese. *Journal of dairy science*, *96*(8), 4816-4829. DOI: 10.3168/jds.2012-6409.
- Carafa I., Stocco G., Nardin T., Larcher R., Bittante G., Tuohy K. & Franciosi E. (2019). Production of naturally γ-aminobutyric acid-enriched cheese using the dairy strains *Streptococcus thermophilus* 84C and *Lactobacillus brevis* DSM 32386. *Frontiers in microbiology*, 10, 93. DOI: 10.3389/fmicb.2019.00093.
- 21. Chia J., Burrow K., Carne A., McConnell M., Samuelsson L., Day L., Young W. & Bekhit A.E.D.A. (2017). Minerals in sheep milk. In R. Watson, R.J. Collier & V. Preedy (Eds.), *Nutrients in Dairy and their Implications on Health and Disease*. (pp. 345-362). Academic Press.
- 22. Cichosz G. & Czeczot H. (2012a). Kwasy tłuszczowe izomerii trans w diecie człowieka. *Bromatologia i Chemia Toksykologiczna*, 45(2), 181-190.
- 23. Cichosz G. & Czeczot H. (2012b). Tłuszcz mlekowy w profilaktyce chorób nowotworowych. *Polski Merkuriusz Lekarski*, *33*(195), 168-172.
- 24. Combarros-Fuertes P., Fernández D., Arenas R., Diezhandino I., Tornadijo M.E. & Fresno J.M. (2015). *Biogenic amines in Zamorano cheese: factors involved in their accumulation. Journal of the Science of Food and Agriculture, 96*(1), 295–305. DOI:10.1002/jsfa.7093.
- 25. Cornu A., Kondjoyan N., Martin B., Verdier-Metz I., Pradel P., Berdagué J.L. & Coulon J.B. (2005). Terpene profiles in Cantal and Saint-Nectaire-type cheese made from raw or pasteurised milk. *Journal of the Science of Food and Agriculture*, 85(12), 2040-2046. DOI: 10.1002/jsfa.2214.
- 26. Costa J.B., de Paula N.T., da Silva P.A., de Souza G.C., Paim A.P.S. & Lavorante A.F. (2019). A spectrophotometric procedure for sialic acid determination in milk employing a flow-batch analysis system with direct heating. *Microchemical Journal*, *147*, 782-788. DOI: 10.1016/j.microc.2019.03.086.
- 27. Creasy W.A., Hankin L. & Handschumaoher R. (1961). Fatty livers induced by orotic acid. 1. Accumulation and metabolism of lipids. *Journal of Biological Chemistry*, 236, 2064-2070.
- 28. Cruz A.G., Faria J.A., Pollonio M.A., Bolini H.M., Celeghini R.M., Granato D. & Shah N.P. (2011). Cheeses with reduced sodium content: Effects on functionality, public health benefits and sensory properties. *Trends in Food Science & Technology*, 22(6), 276-291. DOI: 10.1016/j.tifs.2011.02.003.

- 29. Curioni P.M.G. & Bosset J.O. (2002). Key odorants in various cheese types as determined by gas chromatography-olfactometry. *International Dairy Journal*, *12*(12), 959-984. DOI: 10.1016/S0958-6946(02)00124-3.
- 30. Darewicz M., Borawska J., Minkiewicz P., Iwaniak A. & Starowicz P. (2015). Biologicznie aktywne peptydy uwalniane z białek żywności. Żywność Nauka Technologia Jakość, 3(100), 26-41.
- Demarquoy J., Georges B., Rigault C., Royer M.C., Clairet A., Soty M., Lekounoungou S. & Le Borgne F. (2004). Radioisotopic determination of L-carnitine content in foods commonly eaten in Western countries. *Food Chemistry*, 86(1), 137-142. DOI: 10.1016/j.foodchem.2003.09.023.
- 32. Diana M., Rafecas M., Arco C. & Quilez J. (2014). Free amino acid profile of Spanish artisanal cheeses: Importance of gamma-aminobutyric acid (GABA) and ornithine content. *Journal of Food Composition and Analysis*, *35*, 94-100. DOI: 10.1016/j.jfca.2014.06.007.
- 33. EFSA Panel on Biological Hazards (BIOHAZ) (2011). Scientific opinion on risk based control of biogenic amine formation in fermented foods. *Efsa Journal*, 9(10), 2393.
- 34. Ercan S.S., Bozkurt H. & Soysal Ç. (2013). Significance of biogenic amines in foods and their reduction methods. *Journal of Food Science and Engineering*, 3(8).
- 35. Estrada O., Ariño A. & Juan T. (2019). Salt distribution in raw sheep milk cheese during ripening and the effect on proteolysis and lipolysis. *Foods*, 8(3), 100. DOI: 10.3390/foods8030100/.
- 36. European Commission (2012). Survey on Members States' implementation of the EU salt reduction framework.
- 37. FAOSTAT. (2021). *Livestock Primary and Livestock Processed stats*. Retrieved February 20, 2021, from: <u>http://www.fao.org/faostat/en/#data/QL</u> and *Supply Utilization Accounts*. Retrieved November 14, 2021, from: <u>https://www.fao.org/faostat/en/#data/SCL</u>
- 38. Favaro G., Magno F., Boaretto A., Bailoni L. & Mantovani R. (2005). Traceability of Asiago mountain cheese: a rapid, low-cost analytical procedure for its identification based on solid-phase microextraction. *Journal of dairy science*, 88(10), 3426-3434. DOI: 10.3168/jds.S0022-0302(05)73026-5.
- 39. Fernández D., Arenas R., Gonzalo C., Barbosa E. & Prieto B. (2015). Variation of Fatty Acid Content in Zamorano-Type Ovine Cheese According to the Milk Conjugated Linoleic Acid Content. *Advances in Dairy Research*, *3*(147). DOI: 10.4172/2329-888X.10001.
- Gaglio R., Todaro M., Scatassa M.L., Franciosi E., Corona O., Mancuso I., Di Gerlando R., Cardamone C. & Settanni L. (2019). Transformation of raw ewes' milk applying "Grana" type pressed cheese technology: Development of extra-hard "Gran Ovino" cheese. *International Journal of Food Microbiology*, 307, 108277. DOI: 10.1016/j.ijfoodmicro.2019.108277.
- 41. Gajos E. & Krezlewicz H. (1974). Estimation of orotic acid in milk and milk products. *Przeglad Mleczarski*, 23 (Suppl. 1), 7-9.
- 42. Goa K.L. & Brogden R.N. (1987). *l*-Carnitine. *Drugs 34*, 1-24. DOI: 10.2165/00003495-198734010-00001.
- 43. Gómez-Ruiz J.Á., Ramos M. & Recio I. (2002). Angiotensin-converting enzyme-inhibitory peptides in Manchego cheeses manufactured with different starter cultures. *International Dairy Journal*, *12*(8), 697-706. DOI: 10.1016/s0958-6946(02)0005.
- 44. Gómez-Ruiz J.Á., Taborda G., Amigo L., Recio I. & Ramos M. (2006). Identification of ACE-inhibitory peptides in different Spanish cheeses by tandem mass spectrometry. *European Food Research and Technology*, 223(5), 595-601. DOI: 10.1007/s00217-005-0238-0.
- 45. González-Martín I., Hernández-Hierro J.M., Revilla I., Vivar-Quintana A. & Ortega I.L. (2011). The mineral composition (Ca, P, Mg, K, Na) in cheeses (cow's, ewe's and goat's) with different ripening times using near infrared spectroscopy with a fibre-optic probe. *Food chemistry*, *127*(1), 147-152. DOI: 10.1016/j.foodchem.2010.12.114.
- 46. Govari M., Iliadis S., Papageorgiou D. & Fletouris D. (2020). Seasonal changes in fatty acid and conjugated linoleic acid contents of ovine milk and kefalotyri cheese during ripening. *International Dairy Journal*, 109, 104775. DOI: 10.1016/j.idairyj.2020.104775.
- 47. Gutiérrez-Peña R., Avilés C., Galán-Soldevilla H., Polvillo O., Ruiz Pérez-Cacho P., Guzmán J.L., Horcada A. & Delgado-Pertíñez M. (2021) Physicochemical Composition, Antioxidant Status, Fatty Acid Profile, and Volatile Compounds of Milk and Fresh and Ripened Ewes' Cheese from a Sustainable Part-Time Grazing System. *Foods*, *10*, 80. DOI: 10.3390/foods10010080.
- 48. Horne J., Carpino S., Tuminello L., Rapisarda T., Corallo L. & Licitra G. (2005). Differences in volatiles, and chemical, microbial and sensory characteristics between artisanal and industrial Piacentinu Ennese cheeses. *International Dairy Journal*, *15*(6-9), 605-617. DOI: 10.1016/j.idairyj.2004.10.007.

- 49. Inoue K., Shirai T., Ochiai H., Kasao M., Hayakawa K., Kimura M. & Sansawa H. (2003). Bloodpressure-lowering effect of a novel fermented milk containing gamma-aminobutyric acid (GABA) in mild hypertensives. *European Journal of Clinical Nutrition*, 57(3), 490-495. DOI: 10.1038/sj.ejcn.1601555.
- 50. Irigoyen A., Ortigosa M., Juansaras I., Oneca M. & Torre P. (2007). Influence of an adjunct culture of Lactobacillus on the free amino acids and volatile compounds in a Roncal-type ewe's-milk cheese. *Food Chemistry*, *100*(1), 71-80. DOI: 10.1016/j.foodchem.2005.09.011.
- 51. Jacome-Sosa M.M., Borthwick F., Mangat R., Uwiera R., Reaney M.J., Shen J., Quiroga A.D., Jacobs R.L., Lehner R. & Proctor S.D. (2014). Diets enriched in trans-11 vaccenic acid alleviate ectopic lipid accumulation in a rat model of NAFLD and metabolic syndrome. *The Journal of nutritional biochemistry*, 25(7), 692-701. DOI: 10.1016/j.jnutbio.2014.02.011.
- 52. Janczy A. (2012). Sprzężony kwas linolowy cis-9, trans-11 CLA a zmiany miażdżycowe. Zeszyty Naukowe Akademii Morskiej w Gdyni, (73), 5-15.
- 53. Jansen S.C., van Dusseldorp M., Bottema K.C. & Dubois A.E. (2003). Intolerance to dietary biogenic amines: a review. *Annals of Allergy, Asthma & Immunology, 91*(3), 233-241. DOI: 10.1016/S1081-1206(10)63523-5.
- 54. Jarzynowska A. & Kłopotek, E. (2013). Characteristics of chemical composition and lipid fraction of semi-hard ripening cheese produced from sheep and sheep-cow milk during summer season. *Roczniki* Naukowe Polskiego Towarzystwa Zootechnicznego, 9(4), 39-52.
- 55. Kajimoto O., Hirata H., Nakagawa S., Kajimoto Y., Hayakawa K. & Kimura, M. (2004). Hypotensive effect of fermented milk containing gamma-aminobutyric acid (GABA) in subjects with high normal blood pressure. *Nippon Shokuhin Kagaku Kogaku kaishi*, *51*(2),79-86. DOI: 10.3136/nskkk.51.79.
- 56. Karunanithi D., Radhakrishna A. & Biju V.M. (2013). Quantitative determination of sialic acid in indian milk and milk products. *International Journal of Applied Biotechnology and Pharmaceutical Technology*, 4(1), 318-323.
- 57. Kawęcka A. & Sosin-Bzducha E. (2014). Seasonal changes of the chemical composition of cheese obtained from the milk of indigenous Polish breeds of sheep. *Journal of Animal and Feed Sciences*, 23(2), 131-138. DOI: 10.22358/jafs/65701/2014.
- 58. Kowalska M. & Cichosz G. (2013). Produkty mleczarskie-najlepsze źródło CLA. Bromatologia i Chemia Toksykologiczna, XLVI(1), 1-12.
- 59. Krzęcio-Nieczyporuk E. & Antosik K. (2015). Spożycie wybranych produktów pochodzenia zwierzęcego a zachorowalność na choroby cywilizacyjne. *Przegląd Hodowlany*, 6, 8-11.
- 60. Larson B.L. & Hegarty H.M. (1979). Orotic Acid in Milks of Various Species and Commercial Diary Products. *Journal of Dairy Science*, 62(10), 1641-1644. DOI: 10.3168/jds.S0022-0302(79)83474-8.
- 61. Lim J.N., Oh J.J., Wang T., Lee J.S., Kim S.H., Kim Y.J. & Lee H.G. (2014). Trans-11 18: 1 vaccenic acid (TVA) has a direct anti-carcinogenic effect on MCF-7 human mammary adenocarcinoma cells. *Nutrients*, *6*(2), 627-636. DOI: 10.3390/nu6020627.
- 62. Lima M.R., Bahri H., Morais J.S., Veloso A.C., Fontes L., Lemos E.T. & Peres A.M. (2019). Assessing Serra da Estrela PDO cheeses' origin-production date using fatty acids profiles. *Journal of Food Measurement and Characterization*, 13(4), 2988-2997.
- 63. Linares D.M., O'Callaghan T.F., O'Connor P.M., Ross R.P. & Stanton C. (2016). *Streptococcus thermophilus* APC151 strain is suitable for the manufacture of naturally GABA-enriched bioactive yogurt. *Frontiers in Microbiology*, 7, 1876. DOI: 10.3389/fmicb.2016.01876.
- 64. López-Expósito I., Miralles B., Amigo L. & Hernández-Ledesma B. (2017). Health Effects of Cheese Components with a Focus on Bioactive Peptides. Fermented Foods in Health and Disease Prevention, 239-273. DOI:10.1016/b978-0-12-802309-9.00011-x.
- 65. Malara M., Tkaczyk J., Kęska A., Lutosławska G. & Mazurek K. (2017). Calcium, magnesium and phosphorus dietary intake in active and sedentary Polish students. *Biomedical Human Kinetics*, 9(1), 140-145. DOI: 10.1515/bhk-2017-0020.
- 66. Manolaki P., Katsiari M.C. & Alichanidis E. (2006). Effect of a commercial adjunct culture on organic acid contents of low-fat Feta-type cheese. *Food chemistry*, *98*(4), 658-663. DOI: 10.1016/j.foodchem.2005.06.031.
- 67. Marcone S., Belton O. & Fitzgerald D.J. (2017). Milk-derived bioactive peptides and their health promoting effects: a potential role in atherosclerosis. *British Journal of Clinical Pharmacology*, 83(1), 152-162. DOI: 10.1111/bcp.13002.
- 68. Markiewicz-Kęszycka M., Czyżak-Runowska G., Lipińska P. & Wójtowski J. (2013). Fatty acid profile of milk-a review. *Bulletin of the Veterinary Institute in Pulawy*, *57*(2), 135-139.

- 69. Marrone R., Balestrieri A., Pepe T., Vollano L., Murru N., Michael J.D. & Anastasio A. (2014). Physicochemical composition, fatty acid profile and cholesterol content of "Pecorino Carmasciano" cheese, a traditional Italian dairy product. *Journal of Food Composition and Analysis*, *36*(1-2), 85-89. DOI: 10.1016/j.jfca.2014.05.006.
- Martuscelli M., Gardini F., Torriani S., Mastrocola D., Serio A., Chaves-López C., Schirone M. & Suzzi G. (2005). Production of biogenic amines during the ripening of Pecorino Abruzzese cheese. *International Dairy Journal*, 15(6-9), 571-578. DOI: 10.1016/j.idairyj.2004.11.008.
- 71. Meira S.M.M., Daroit D.J., Helfer V.E., Corrêa A.P.F., Segalin J., Carro S. & Brandelli A. (2012). Bioactive peptides in water-soluble extracts of ovine cheeses from Southern Brazil and Uruguay. *Food Research International*, 48(1), 322-329. DOI: 10.1016/j.foodres.2012.05.009.
- 72. Mikami Y. (1988). Microbial conversion of terpenoids. *Biotechnology and genetic engineering reviews*, 6(1), 271-320. DOI: 10.1080/02648725.1988.10647850.
- 73. Milewski S. (2006). Walory prozdrowotne produktów owczych. *Medycyna Weterynaryjna*, 62(5), 516-519.
- 74. Milewski S., Ząbek K., Antoszkiewicz Z., Tański Z. & Błażejak J. (2016). Walory prozdrowotne serów z mleka owczego i koziego wytworzonych w gospodarstwach Warmii i Mazur. *Przegląd Hodowlany*, 2, 20-22.
- 75. Miller A., McGrath E., Stanton C. & Devery R. (2003). Vaccenic acid (t11–18: 1) is converted to c9, t11-CLA in MCF-7 and SW480 cancer cells. *Lipids*, *38*(6), 623-632. DOI: 10.1007/s11745-003-1107-8.
- 76. Miner-Williams W.M., Stevens B.R. & Moughan P.J. (2014). Are intact peptides absorbed from the healthy gut in the adult human? *Nutrition research reviews*, 27(2), 308-329. DOI: 10.1017/S0954422414000225.
- 77. Moran L., Aldezabal A., Aldai N. & Barron L.J.R. (2019). Terpenoid traceability of commercial sheep cheeses produced in mountain and valley farms: From pasture to mature cheeses. *Food Research International*, *126*, 108669. DOI: 10.1016/j.foodres.2019.108669.
- 78. Muñoz N., Ortigosa M., Torre P. & Izco J.M. (2003). Free amino acids and volatile compounds in an ewe's milk cheese as affected by seasonal and cheese-making plant variations. *Food Chemistry*, 83(3), 329-338. DOI: 10.1016/S0308-8146(03)00133-X.
- 79. Murgia M.A., Deiana P., Nudda A., Correddu F., Montanari L. & Mangia N.P. (2020). Assessment of microbiological quality and physicochemical parameters of Fruhe made by ovine and goat milk: A Sardinian (Italy) cheese. *Fermentation*, 6(4), 119. DOI: 10.3390/fermentation6040119.
- Murru E., Carta G., Cordeddu L., Melis M.P., Desogus E., Ansar H., Chilliard Y., Ferlay A., Stanton C., Coakley M., Ross R.P., Piredda G., Addis M., Mele M.C., Cannelli G., Banni S. & Manca, C. (2018). Dietary Conjugated Linoleic Acid-Enriched Cheeses Influence the Levels of Circulating n-3 Highly Unsaturated Fatty Acids in Humans. *International Journal of Molecular Sciences*. 19(6), 1730. DOI: 10.3390/ijms19061730.
- 81. Nagao K. & Yanagita T. (2010). Medium-chain fatty acids: functional lipids for the prevention and treatment of the metabolic syndrome. *Pharmacological Research*, 61(3), 208-212. DOI: 10.1016/j.phrs.2009.11.007.
- 82. Nakano T., Sugawara M. & Kawakami H. (2001). Sialic acid in human milk: composition and functions. *Acta Paediatr Taiwan*, 42(1), 11-17.
- Nomura M., Kimoto H., Someya Y., Furukawa S. & Suzuki I. (1998). Production of γ-aminobutyric acid by cheese starters during cheese ripening. *Journal of Dairy Science*, 81(6), 1486-1491. DOI: 10.3168/jds.S0022-0302(98)75714-5.
- 84. Nudda A., McGuire M.A., Battacone G. & Pulina G. (2005). Seasonal variation in conjugated linoleic acid and vaccenic acid in milk fat of sheep and its transfer to cheese and ricotta. *Journal of Dairy Science*, 88(4), 1311-1319. DOI: 10.3168/jds.S0022-0302(05)72797-1.
- 85. OECD/FAO. (2021). *OECD-FAO Agricultural Outlook 2020-2029*. Retrieved November 14, 2021, from <u>https://www.oecd-ilibrary.org/agriculture-and-food/cheese-projections-consumption-food_leddd347-en</u>
- 86. Ordonez A.I., Ibanez F.C., Torre P. & Barcina Y. (1997). Formation of biogenic amines in Idiazábal ewe's-milk cheese: effect of ripening, pasteurization, and starter. *Journal of Food Protection*, 60(11), 1371-1375. DOI: 10.4315/0362-028X-60.11.1371.
- 87. Oruch R. & Pryme I.F. (2012). The biological significance of vitamin A in humans: A review of nutritional aspects and clinical considerations. *Science Jet*, *1*(19), 1-13.

- 88. Papademas P. & Robinson R.K. (2002). Some Volatile Plant Compounds in Halloumi Cheeses made from Ovine or Bovine Milk. *LWT Food Science and Technology*, *35*(6), 512–516. DOI: 10.1006/fstl.2002.0901.
- 89. Pariza M.W. (1999). The biological activities of conjugated linoleic acid. In M.P. Yurawecz, M.M. Mossoba, J.K.G. Kramer, M.W. Pariza & G.J. Nelson (Eds.), *Advances in conjugated linoleic acid research*, Vol. 1. (pp. 12-20), AOCS Press.
- 90. Partidário A.M., Ribeiro J.C. & Prates J.A. (2008). Fatty acid composition and nutritional value of fat in three PDO ewe's milk Portuguese cheeses. *Dairy Science and Technology*, 88(6), 683-694. DOI: 10.1051/dst:2008032.
- Pękala J., Patkowska-Sokoła B., Bodkowski R., Jamroz D., Nowakowski P., Lochynski S. & Librowski, T. (2011). L-carnitine-metabolic functions and meaning in humans life. *Current Drug Metabolism*, 12(7), 667-678. DOI: 10.2174/138920011796504536.
- 92. Perretti G., Marconi O., Montanari L. & Fantozzi, P. (2004). Rapid determination of total fats and fatsoluble vitamins in Parmigiano cheese and salami by SFE. *LWT-Food Science and Technology*, *37*(1), 87-92. DOI: 10.1016/S0023-6438(03)00138-5.
- 93. Pinho O., Ferreira I.M., Mendes E., Oliveira B.M. & Ferreira M. (2001). Effect of temperature on evolution of free amino acid and biogenic amine contents during storage of Azeitão cheese. *Food Chemistry*, 75(3), 287-291. DOI: 10.1016/S0308-8146(01)00109-1.
- 94. Pintado A.I., Pinho O., Ferreira I.M., Pintado M.M.E., Gomes A.M. & Malcata F.X. (2008). Microbiological, biochemical and biogenic amine profiles of Terrincho cheese manufactured in several dairy farms. *International Dairy Journal*, *18*(6), 631-640. DOI: 10.1016/j.idairyj.2007.11.021.
- 95. Pintus S., Murru E., Carta G., Cordeddu L., Batetta B., Accossu S., Pistis D., Uda S., Elena Ghiani M., Mele M., Secchiari P., Almerighi G., Pintus P. & Banni, S. (2013). Sheep cheese naturally enriched in α-linolenic, conjugated linoleic and vaccenic acids improves the lipid profile and reduces anandamide in the plasma of hypercholesterolaemic subjects. *British Journal of Nutrition*, 109(8), 1453-1462. DOI: 10.1017/S0007114512003224.
- 96. Pisanu S., Pagnozzi D., Pes M., Pirisi A., Roggio T., Uzzau S. & Addis M.F. (2015). Differences in the peptide profile of raw and pasteurised ovine milk cheese and implications for its bioactive potential. *International Dairy Journal*, *42*, 26-33. DOI: 10.1016/j.idairyj.2014.10.007.
- 97. Poulopoulou I., Zoidis E., Massouras T. & Hadjigeorgiou I. (2011). Terpenes transfer to milk and cheese after oral administration to sheep fed indoors. *Journal of Animal Physiology and Animal Nutrition*, 96(2), 172–181. DOI: 10.1111/j.1439-0396.2011.0.
- 98. Poveda J.M., Chicón R., & Cabezas L. (2015). Biogenic amine content and proteolysis in Manchego cheese manufactured with Lactobacillus paracasei subsp. paracasei as adjunct and other autochthonous strains as starters. *International Dairy Journal*, 47, 94-101. DOI: 10.1016/j.idairyj.2015.03.004.
- 99. Prandini A., Sigolo S. & Piva G. (2011). A comparative study of fatty acid composition and CLA concentration in commercial cheeses. *Journal of Food Composition and Analysis*, 24(1), 55-61. DOI: 10.1016/j.jfca.2010.04.004.
- 100. Prandini A., Sigolo S., Tansini G., Brogna N. & Piva G. (2007). Different level of conjugated linoleic acid (CLA) in dairy products from Italy. *Journal of Food Composition and Analysis*, 20(6), 472–479. DOI: 10.1016/j.jfca.2007.03.001.
- 101. Ptasińska-Marcinkiewicz J. (2014). Hodowla owiec i produkcja mleka owczego w Polsce i na świecie. Zeszyty Naukowe Uniwersytetu Ekonomicznego w Krakowie, 3(927), 43-55.
- 102. Rashidinejad A., Bremer P., Birch, J. & Oey, I. (2017). Nutrients in Cheese and Their Effect on Health and Disease. Academic Press.
- 103. Raynal-Ljutovac K., Lagriffoul G., Paccard P., Guillet I. & Chilliard Y. (2008). Composition of goat and sheep milk products: An update. *Small ruminant research*, 79(1), 57-72. DOI: 10.1016/j.smallrumres.2008.07.009.
- Recio I., de la Fuente M.A., Juárez M. & Ramos M. (2009). Bioactive components in sheep milk. In Y. W. Park (Ed.), *Bioactive components in milk and dairy products*. (pp. 83-104), John Wiley & Sons.
- 105. Redruello B., Saidi Y., Sampedro L., Ladero V., Del Rio B. & Alvarez M.A. (2021). GABA-Producing Lactococcus lactis Strains Isolated from Camel's Milk as Starters for the Production of GABA-Enriched Cheese. *Foods*, *10*(3), 633. DOI: 10.3390/foods10030633.
- 106. Renes E., Ladero V., Tornadijo M.E. & Fresno J.M. (2019). Production of sheep milk cheese with high γ-aminobutyric acid and ornithine concentration and with reduced biogenic amines level using autochthonous lactic acid bacteria strains. *Food microbiology*, 78, 1-10. DOI: 10.1016/j.fm.2018.09.003.

- 107. Revilla I., Lobos Ortega I.A., Vivar-Quintana A.M., González-Martín M.I., Hernández Hierro J.M. & González Pérez C. (2014). Variations in the contents of vitamins A and E during the ripening of cheeses with different compositions. *Czech Journal of Food Sciences*, *32*(4), 342-347. DOI: 10.13140/2.1.2177.0884.
- 108. Rizvi S., Raza S.T., Ahmed F., Ahmad A., Abbas S. & Mahdi F. (2014). The role of vitamin e in human health and some diseases. *Sultan Qaboos University Medical Journal*, *14*(2), e157-65.
- 109. Rizzello C.G., Losito I., Gobbetti M., Carbonara T., De Bari M.D. & Zambonin P.G. (2005). Antibacterial activities of peptides from the water-soluble extracts of Italian cheese varieties. *Journal of Dairy Science*, 88(7), 2348-2360. DOI: 10.3168/jds.S0022-0302(05)72913-1.
- 110. Robinson R.K. & Tamime A.Y. (1996). Feta & Related Cheeses. CRC Press.
- 111. Roche H.M., Noone E. & Gibney A.N.M.J. (2001). Conjugated linoleic acid: a novel therapeutic nutrient? *Nutrition Research Reviews*, *14*(1), 173-188. DOI: 10.1079/NRR200122.
- 112. Rospond B. & Chłopicka J. (2013). Funkcje biologiczne L-karnityny i jej zawartość w wybranych produktach spożywczych. *Przegląd Lekarski*, 70(2), 85-91.
- 113. Rutkowska E., Tambor K., Rutkowska J. & Stołyhwo A. (2015). Charakterystyka prozdrowotnych kwasów tłuszczowych tłuszczu mlecznego. *Problemy Higieny i Epidemiologii*, *96*(2), 377-386.
- 114. Salque M., Bogucki P.I., Pyzel J., Sobkowiak-Tabaka I., Grygiel R., Szmyt M. & Evershed R.P. (2013). Earliest evidence for cheese making in the sixth millennium BC in northern Europe. *Nature*, 493(7433), 522. DOI: 10.1038/nature11698.
- 115. Samraj A.N., Pearce O.M., Läubli H., Crittenden A.N., Bergfeld A.K., Banda K., Gregg C.J., Bingman A.E., Secrest P., Diaz S.L., Varki N.M. & Varki A. (2015). A red meat-derived glycan promotes inflammation and cancer progression. *Proceedings of the National Academy of Sciences of the United States of America*, *112*(2), 542-547. DOI: 10.1073/pnas.1417508112.
- 116. Sanjuán E., Saavedra P., Millán R., Castelo M. & Fernández-Salguero J. (1998). Effect of ripening and type of rennet on the mineral content of Los Pedroches cheese. *Journal of food quality*, 21(3), 187-200. DOI: 10.1111/j.1745-4557.1998.tb00515.x.
- 117. Santiago-López L., Aguilar-Toalá J.E., Hernández-Mendoza A., Vallejo-Cordoba B., Liceaga A.M. & González-Córdova A.F. (2018). Invited review: Bioactive compounds produced during cheese ripening and health effects associated with aged cheese consumption. *Journal of dairy science*, 101(5), 3742-3757. DOI: 10.3168/jds.2017-13465.
- 118. Santos M.S. (1996). Biogenic amines: their importance in foods. International Journal of Food Microbiology, 29(2-3), 213-231. DOI: 10.1016/0168-1605(95)00032-1.
- 119. Sauer L.A., Dauchy R.T., Blask D.E., Krause J.A., Davidson L.K., Dauchy E.M., Welham K.M. & Coupland K. (2004). Conjugated linoleic acid isomers and trans fatty acids inhibit fatty acid transport in hepatoma 7288CTC and inguinal fat pads in Buffalo rats. *The Journal of nutrition*, *134*(8), 1989-1997. DOI: 10.1093/jn/134.8.1989.
- Schirone M., Tofalo R., Fasoli G., Perpetuini G., Corsetti A., Manetta A.C., Ciarrocchi A. & Suzzi G. (2013). High content of biogenic amines in Pecorino cheeses. *Food microbiology*, 34(1), 137-144. DOI: 10.1016/j.fm.2012.11.022.
- 121. Schirone M., Tofalo R., Mazzone G., Corsetti A. & Suzzi, G. (2011). Biogenic amine content and microbiological profile of Pecorino di Farindola cheese. *Food Microbiology*, 28(1), 128-136. DOI: 10.1016/j.fm.2010.09.005
- 122. Schirone M., Tofalo R., Visciano P., Corsetti A., Suzzi G. (2012). Biogenic amines in Italian Pecorino cheese. *Frontiers in microbiology*, *3*, 171. DOI: 10.3389/fmicb.2012.00171.
- 123. Seline K.G. & Johein H. (2007). The determination of L-carnitine in several food samples. *Food chemistry*, 105(2), 793-804. DOI: 10.1016/j.foodchem.2007.01.058.
- 124. Siragusa S., De Angelis M., Di Cagno R., Rizzello C.G., Coda R. & Gobbetti M. (2007). Synthesis of γ-aminobutyric acid by lactic acid bacteria isolated from a variety of Italian cheeses. *Applied and Environmental Microbiology*, 73(22), 7283-7290. DOI: 10.1128/AEM.01064-07.
- 125. Sofi F., Buccioni A., Cesari F., Gori A.M., Minieri S., Mannini L., Casini A., Gensini G.F., Abbate R. & Antongiovanni M. (2010). Effects of a dairy product (pecorino cheese) naturally rich in cis-9, trans-11 conjugated linoleic acid on lipid, inflammatory and haemorheological variables: a dietary intervention study. *Nutrition, Metabolism & Cardiovascular Diseases*, 20(2), 117-124. DOI: 10.1016/j.numecd.2009.03.004.
- 126. Szczurek W. (2004). Kwas sjalowy nowe spojrzenie na wpływ produktów pochodzenia zwierzęcego na organizm człowieka. *Wiadomości Zootechniczne*, 42(4), 27-36.

- 127. Tavaria F.K., Franco I., Carballo F.J. & Malcata F.X. (2003). Amino acid and soluble nitrogen evolution throughout ripening of Serra da Estrela cheese. *International Dairy Journal*, *13*(7), 537-545. DOI: 10.1016/S0958-6946(03)00060-8.
- 128. Tofalo R., Perpetuini G., Battistelli N., Pepe A., Ianni A., Martino G. & Suzzi, G. (2019). Accumulation γ-Aminobutyric Acid and Biogenic Amines in a Traditional Raw Milk Ewe's Cheese. *Foods*, 8(9), 401. DOI: 10.3390/foods8090401.
- 129. Tong X., Chen G.C., Zhang Z., Wei Y.L., Xu J.Y. & Qin L.Q. (2017). Cheese Consumption and Risk of All-Cause Mortality: A Meta-Analysis of Prospective Studies. *Nutrients*, 9(1), 63. DOI: 10.3390/nu9010063.
- 130. Torracca, B., Nuvoloni, R., Ducci, M., Bacci, C., & Pedonese, F. (2015). Biogenic amines content of four types of "Pecorino" cheese manufactured in Tuscany. *International Journal of Food Properties*, *18*(5), 999-1005.
- 131. Urbienė, S. & Leskauskaitė, D. (2006). Formation of some organic acids during fermentation of milk. *Polish Journal of Food and Nutrition Sciences*, 15(3), 277-281.
- 132. Valdivielso I., de Renobales M., Aldai N. & Barron L.J.R. (2017). Changes in terpenoid composition of milk and cheese from commercial sheep flocks associated with seasonal feeding regimens throughout lactation. *Journal of dairy science*, *100*(1), 96-105. DOI: 10.3168/jds.2016-11761.
- 133. Valsamaki K., Michaelidou A., & Polychroniadou A. (2000). Biogenic amine production in Feta cheese. *Food chemistry*, 71(2), 259-266. DOI: 10.1016/S0308-8146(00)00168-0.
- 134. Verzera A., Condurso C., Ziino, M., Romeo V., Todaro M., Conte F. & Dima G. (2010). Free fatty acids and other volatile compounds for the characterisation of "Vastedda della valle del Belice" cheese Acidos grasos libres y otros constituyentes volátiles para la caracterización de queso "Vastedda della vella del Belice". *CyTA–Journal of Food*, 8(3), 237-243. DOI: 10.1080/19476330903450282.
- 135. Walther B., Schmid A., Sieber R. & Wehrmüller, K. (2008). Cheese in nutrition and health. *Dairy Science & Technology*, 88(4-5), 389-405. DOI: 10.1051/dst:2008012.
- 136. Wang B. (2009). Sialic acid is an essential nutrient for brain development and cognition. *Annual Review* of Nutrition, 29, 177-222. DOI: 10.1146/annurev.nutr.28.061807.155515.
- 137. Watanabe M., Maemura K., Kanbara K., Tamayama T. & Hayasaki, H. (2002). GABA and GABA receptors in the central nervous system and other organs. *International review of cytology*, *213*, 1-47. DOI: 10.1016/s0074-7696(02)13011-7.
- 138. West T.P., Chunduru J. & Murahari E.C. (2017). Orotic Acid: Why it is Important to Understand Its Role in Metabolism. *Biochemistry & Physiology*, 6(1), 1000e157. DOI: 10.4172/2168-9652.1000e157.
- 139. Woollard D.C., Indyk H.E. & Woollard, G.A. (1999). Carnitine in milk: a survey of content, distribution and temporal variation. *Food chemistry*, 66(1), 121-127. DOI: 10.1016/S0308-8146(99)00042-4.
- 140. Wright L.D., Huff J.W., Skeggs H.R., Valentik K.A. & Bosshardt, D.K. (1950). Orotic acid, a growth factor for *Lactobacillus bulgaricus*. *Journal of the American Chemical Society*, 72(5), 2312-2313. DOI: 10.1021/ja01161a544.
- 141. Yoto A., Murao S., Motoki M., Yokoyama Y., Horie N., Takeshima K., Masuda K., Kim M. & Yokogoshi H. (2012). Oral intake of γ -aminobutyric acid affects mood and activities of central nervous system during stressed condition induced by mental tasks. *Amino Acids*, 43(3), 1331-1337. DOI: 10.1007/s00726-011-1206-6.
- 142. Zamberlin Š., Antunac N., Havranek J. & Samaržija D. (2012). Mineral elements in milk and dairy products. *Mljekarstvo*, 62(2), 111-125.
- 143. Zazzu C., Addis M., Caredda M., Scintu M.F., Piredda G. & Sanna G. (2019). Biogenic amines in traditional fiore Sardo PDO sheep cheese: Assessment, validation and application of an RP-HPLC-DAD-UV method. *Separations*, 6(1), 11. DOI: 10.3390/separations6010011.
- 144. Zdrojewicz Z., Zyskowska K. & Górecka A. 2017. Wpływ substancji zawartych w serach na organizm człowieka. *Medycyna Rodzinna*, 20(2), 124-129.
- 145. Živný P., Živná H., Pavlíková L., Hrubá P., PaličkaV., Soukup T. & Šimáková, E. (2007). The effect of cholesterol and orotic acid administration and methionin-cholin deficiency on liver DNA synthesis and lipid metabolism in rats. *Folia Gastroenterologica et Hepatologica*, *5*, 3-4.
- 146. Zlatanos S., Laskaridis K., Feist C. & Sagredos A. (2002). CLA content and fatty acid composition of Greek Feta and hard cheeses. *Food Chemistry*, 78(4), 471-477. DOI: 10.1016/S0308-8146(02)00159-0.